10.1053.paor.2000.0265 available online at http://www.idealibrary.com on IDEAL

## **ARTICLE**

# Detection and Isolation of MUC1 Mucin from Larynx Squamous Cell Carcinoma

María V CROCE, Mike R PRICE, 1 Amada SEGAL-EIRAS

Centro de Investigaciones Inmunológicas Básicas y Aplicadas (CINIBA), Universidad Nacional de la Plata, Argentina; 
<sup>1</sup>Cancer Research Laboratory, School of Pharmaceutical Sciences, University of Nottingham, United Kingdom

The progression from uncontrolled cell proliferation to invasion and metastasis of epithelial tumors is partially understood. Alteration of epithelial mucin expression have been described in different malignant localizations but only few attempts have been made to identify mucin expression in malignant laryngeal tumors. In the present report, results are shown of studies on the expression of mucins and carbohydrate related antigens in laryngeal cancer and on the isolation of MUC1 mucin from this tumor tissue. Malignant laryngeal specimens were processed for immunohistochemical analysis and for extranuclear membrane fractions (ENM) which were obtained by ultracentrifugation. Subsequently, ENM samples were centrifuged in density-gradient; the analysis of fractions was performed by means of SDS-PAGE and Western-blotting. The panel of monoclonal antibodies (MAbs) included anti MUC1 mucin, anti Lewis x, anti sialyl Lewis x, anti Lewis y, anti MUC-5B, anti oral mucin (gp230), anti Tn hapten, anti p53 and anti cytokeratins. By immunohistochemistry, it was possible to detect MUC1 mucin, Lewis x and Lewis y showing strong reactions while sialyl-Lewis x and Tn antigen only reacted weakly in a few cells; cytokeratins were detected in all samples. In ENM derived fractions obtained by CsCl centrifugation, MUC1 was demonstrated by Western blotting. Conclusions: (1) laryngeal cancer antigenic expression comprises mostly MUC1 mucin, Lewis x, Lewis y as well as Tn antigen and (2) the methodology here employed is useful to isolate MUC1 from tumor samples. (Pathology Oncology Research Vol 6, No 2, 93-99, 2000)

Keywords: laryngeal carcinoma, tumor antigens, MUC1mucin

## Introduction

Worldwide, incidence of larynx tumors are on increase, <sup>22,31</sup> smoking of cigarettes and/or alcohol consumption have been etiologically associated with their carcinogenesis. Different antigenic changes occur within epithelial tumor development; among these, mucins may be altered during the neoplastic process and progression to a metastatic phenotype. <sup>16</sup>

These molecules are produced and secreted by a wide variety of normal and neoplastic epithelial cells; during the last decade, different mucins associated to carcinoma have been investigated being described at least nine human mucin genes (MUC1-4, MUC5B, MUC5AC,

Received: March 17, 2000; accepted: April 10, 2000 Correspondence: Professor Amada SEGAL-EIRAS, CINIBA, Calle 60 y 120 Universidad Nacional La Plata, Argentina. Fax 0054 221 4259115; E-mail: as-eiras@netverk.com.ar MUC6-8). <sup>13,42</sup> MUC1 mucins are high molecular weight transmembrane glycoproteins which extracellular domain is made up of tandem repeats of 20 aminoacids characterized by many O-linked oligosaccharides attached to the mucin core polypeptides through serine and threonine residues. <sup>12</sup> The complexity of carbohydrate moieties have been investigated proving the presence of truncated side chains in carcinoma as result of changes in glycosyl and sialyltransferases. <sup>47</sup> Mucin alterations have been related with cellular motility, and consequently, with tumor invasion and metastatic dissemination; <sup>41,36</sup> on the other hand, several tumor antigens have been detected related to tumor mucin molecules which may induce cellular <sup>2,19,21,38</sup> and humoral immune responses in the host. <sup>7,23</sup>.

Although progress has been made in the characterization of mucin structure and organ specific expression, few attempts were undertaken to identify mucin expression in malignant tumors of the larynx.

94 CROCE et al

In the present report we discuss results obtained on the expression of mucin antigens as well as the MUC1 mucin isolation from laryngeal carcinoma cells.

### Materials and Methods

## Tissue samples

The present study was performed in a total of 20 malignant laryngeal tumors belonging to patients treated with surgical resection and classified according to UICC criteria TNM Classification of Malignant Tumors, 5<sup>th</sup> edition. <sup>18</sup> Clinicopathological characteristics of patients are depicted in *Table 1*.

Normal squamous epithelia from the same localization were obtained by biopsy and they were included as controls.

Tissues were processed following routine directives; specimens were sectioned to cover different programmed studies (*Figure 1*); a piece of tissue was fixed in formaldehyde for histopathological diagnosis and immunohistochemical analysis while another tissue sample was rinsed with fresh sterile Hank's balanced cell solution and subsequent processed for the preparation of subcellular fractions.

All human studies have been reviewed by the University of La Plata Ethical Committee and experiments were performed according with the Helsinki Declaration. Informed consent was obtained from all patients included in this study.

## Preparation of extranuclear membrane fractions

Fractions were prepared from human tumor tissues.<sup>32</sup> Briefly, tissues were homogeneized in 1.41M PBS pH 7.2, at 4ml/g; homogenates were centrifuged at 600 x g and at 105000 x g at 4°C and precipitates (extranuclear membrane fraction) were resuspended in 1.41M PBS, liophylized and stored at -20°C for subsequent density gradient centrifugation.

## Density-gradient centrifugation

Samples from extranuclear membrane fractions in 4 M guanidium chloride 1% NP40 were centrifuged in 6 M guanidium chloride/CsCl (density 1.45 g/ml) at 40000 rpm for 66 h at 10°C in a Beckman Titanium 70.1 fixed-angle rotor; 500  $\mu$ l fractions were collected and the density of each fraction determined using a Hamilton syringe (Merck, Dagenham, Essex, UK) as a pycnometer. 45

## SDS-PAGE and Immunoblotting

Fractions were collected and dialysed against 1.41 M PBS at 4°C for 48 h and then liophylized, resuspended in SDS-PAGE sample buffer at reducing conditions and run following standard procedures<sup>24</sup> in a discontinuous buffer

system. After electrophoresis gels were either stained with Coomassie blue or they were transferred electrophoretically to nitrocellulose membranes<sup>46</sup> and incubated with different monoclonal antibodies.

### Immunohistochemical analysis

This technique was developed according to a previous report, with minor modifications. All specimens were fixed in phosphate buffered formalin, embedded in paraffin and cut into 5  $\mu$ m serial sections. Deparaffinized sections were treated with 10mM sodium citrate buffer at 100°C for 5 minutes, then, they were incubated overnight at 4°C with mouse monoclonal antibodies. Negative controls were incubated with PBS instead of monoclonal antibodies.

The whole area of each sample was observed by sequentially examining low power (x10) optical fields; the staining of cytoplasm, plasma and nuclear membranes were also evaluated. Cells were considered positive when at least one of these components was stained; heterogeneity was graded according to positive reactions, intensity and distribution. Staining intensity was graded as negative, low, moderate and strong.<sup>11</sup>

With respect to p53, and considering other reports,<sup>34</sup> a sample was coded as positive even when only a few cells were reactive.

Table 1 Clinicopathological characteristics of patients.

Characteristics	No. of patients (%)				
Sex					
Male	20/20				
Female	0/20				
Age, years					
<65	2/20				
>65	18/20				
T stage					
T1	0/20				
T2	2/20				
T3	9/20				
T4	9/20				
N stage					
N0	0/20				
N1	4/20				
N2	16/20				
Metastases					
M0	19/20				
M1	1/20				
Differentiation					
Well differentiated	2/20				
Moderately differentiated	9/20				
Poorly differentiated	9/20				

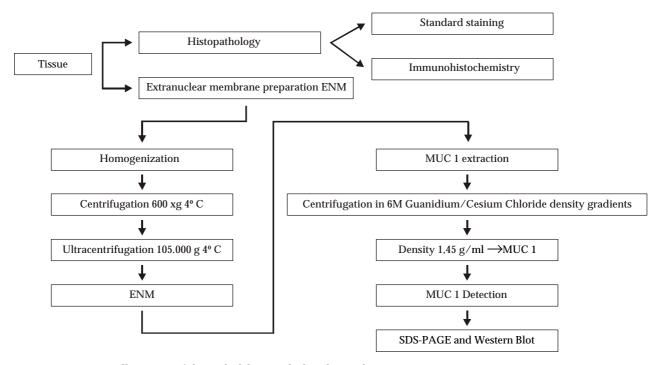


Figure 1. Summary illustration of the methodology applied in this work.

## Monoclonal antibodies (Mabs)

The following monoclonal antibodies (Mabs) were assayed: C14 Mab, an IgM anti-Lewis y hapten against the difucosylated Type-2 blood group chains,4 Mab KM93 (IgM), an anti syalil-Lewis x and KM380 (IgM), an anti Lewis x. 15 Three anti MUC1 Mabs were employed: Mab C-595 (IgG3) defines the tetrameric epitope Arg-Pro-Ala-Pro in the MUC1 protein core, 32 HMFG1 and HMFG243 define the epitopes PDTR and DTR respectively.<sup>33</sup> Mab 83D4 (IgM) against Tn determinant,<sup>30</sup> an anti MUC-5B (PANH2 Mab) (IgG1) raised against partially deglycosylated MG1,28 PANH3, an anti MUC-7 and PANH4 Mab (IgM) an anti gp230 derived from buccal mucosa.<sup>27</sup> Also, anti-p53 Mab (mouse IgG2a isotype) (SIGMA No. P-5813) and finally, an anti-human cytokeratin (CK)/HRP against keratins 5, 6, 8, 17 and 19 (Dako EPOS, Code No. U 7022) was assayed.

### Results

By routine techniques, histopathological examination was performed; all tumors were identified as invasive squamous cell carcinoma of the larynx; it was possible to observe two different types of cells: tumor large cells and small ones with intense staining and with round nuclei; prominent anaplasia, anisocytosis as well as anisocariosis were present; bizarre mitotic figures were also seen. In a group of samples, the expression of different tumor anti-

gens was studied using a panel of monoclonal antibodies; results are summarised in *Table 2*.

MUC1 mucin was detected with C595 MAb in four out of seven tumor patients' samples; in most reactive cells, a homogeneous cytoplasmic immune-staining was shown (*Figure 2a*); often, a perinuclear reaction was detected; also several cellular membranes revealed a positive staining while nuclei remained negative the other anti-MUC1 MAbs did not display any reactivity.

Another feature of the distribution of antigenic expression was a strong cytoplasmic reaction observed with anti Lewis x MAb (KM380) in four out of seven samples while only a moderate reaction was detected in one sample; an example of one tumor is depicted in *Figure 2b*. C14 MAb (anti Lewis y antigen) showed a consistent staining restricted to the cytoplasm in some cells; 83D4 (antiTn hapten) stained weakly with a homogeneous pattern while KM93 MAb (anti-sialyl Lewis x) only reacted in a few cells in some tumor specimens.

All samples stained with the anticytokeratin MAb mostly in the cytoplasm while p53 was widely detected in cell nucleus; only one sample showed a positive reaction with anti-oral mucin gp230 while negative results were obtained with the other MAbs: anti-MUC-5B as well as MUC-7.

Normal squamous epithelia belonging to the same localization were analysed; expression of oral mucin (gp230), and Lewis x was found in three out of seven samples considered; sialyl Lewis x and Lewis y in 4/7 while MUC1

96 CROCE et al

Table 2. Summary of i	immunohistochemical	analysis of tumor	samples
-----------------------	---------------------	-------------------	---------

Patient	ient Monoclonal Antibodies									
Number	Lewis y	Sialyl Lewis x	Lewis x	MUC1	Tn hapten	Muc-5B	Muc-7	Gp230	P53	Cytokeratins
1	G. C.	¢.	••	•	C: C	0	0	0	Ç: C	C: C
2	Ĉ	0	••	0	2	0	0	0	•	8
3	Ĉ	8	•	0	8	0	0	0	o o	6
4	0	0	0	•	0	0	0	Ĉ.	•	6.6
5	0	0	0	0	0	0	0	0	Ĉ.	•
6	0	0	••	0	8	0	0	0	Ĉ.	•
7	8	<b>C</b> . <b>C</b> .	••	©.	0	0	0	0	•	•

O = negative

a = mild positive reaction

• = moderate positive reaction

● ● = strong positive reaction

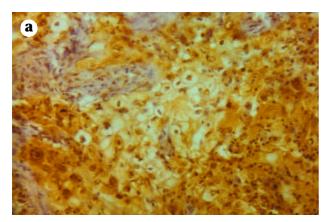
and Tn antigen in 2/7. The outstanding feature of normal expression was its restriction to a few cells belonging to one epithelial stratum being the pattern of distribution quite homogeneous; mainly a weak cytoplasmic reaction was detected in all samples with one exception: MUC1 was exclusively expressed at cellular membranes. In one specimen, sially Lewis x was expressed in a few cells belonging to different epithelial strata; on the other hand, cytokeratins were detected in small amounts with a weak and homogeneous cytoplasmic pattern while in most cases, p53 was observed in a few cells from one normal specimen.

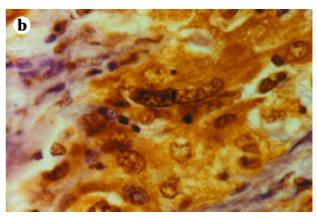
In order to isolate MUC1 mucin from laryngeal tumors, extranuclear membranes were prepared and analysed by SDS-PAGE and Western-blotting incubated with anti MUC1 Mabs. In specimen belonging to patient number 1, these studies showed a positive reaction with C595 Mab; the same positive staining was obtained with anti Lewis x and anti Tn hapten of the same tumor sample (data not

shown). Extranuclear membranes isolated from this tumor sample were subjected to density gradient centrifugation in guanidin Cs Cl; fractions obtained were subsequently analysed by SDS-PAGE and Western-blotting employing the panel of MAbs. Fraction 7 (from a density of 1.43 g/ml), reacted with anti MUC1 C595 Mab by Western-blot; the major feature of staining pattern was the presence of two bands of high-molecular-mass material of more than 180 kDa and some trace minor bands (*Figure 3*); this fraction also showed reactivity with anti Lewis x and anti Tn hapten employing the same technique.

## Discussion

Neoplastic cells express a wide variety of molecules since the development and progression of cancer are regulated by the expression of different molecules encoded by a continuous enlarged list of diverse oncogenes and suppressor genes. Most immuno-biological functions as well



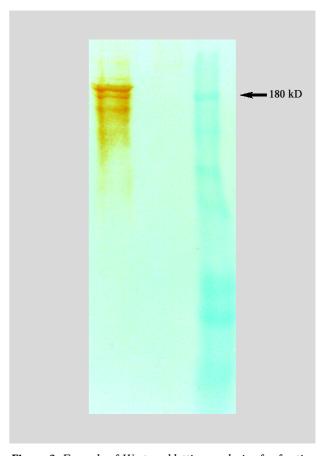


**Figure 2. a)** Immunohistochemistry of larynx carcinoma with anti MUC1 MAb C595. Intense staining is observed in cytoplasma and plasmatic membrane. Non-reactive cells show a clear cytoplasm. (x100). **b)** Immunereactivity obtained by incubation with antiLewis x Mab (KM380). The positive reaction is observed in the cytoplasm with a vesicle pattern; mitosis in the centre of the picture (x630).

as clinical properties of these molecules are still unknown, although some progress has been obtained in recent years. This relationship should be useful for the identification of accurate indicators in diverse aspects of neoplastic phenomena from basic knowledge to clinical practice.

Mucin alteration occurs during tumor development, such changes include differences in mucin-associated antigenic expression between cancer cells and their normal counterparts. By immunohistochemistry, series of MAbs against epithelial mucins and carbohydrate related antigens have been evaluated. It was possible to show a high expression of mucins and/or carbohydrate related antigens such as Lewis x, Lewis y and Tn hapten being sialyl Lewis x also expressed in a small number of samples; furthermore, this expression may be related to an advanced malignant disease since most tumors included in the present study were disseminated. Tumor associated antigens here evaluated have been reported to be factors associated with increased metastatic potential; 16,35,36 for instance, diverse mechanisms have shown to be involved in MUC1 cellular adhesion to vascular endothelium through exposing usually cryptic associated carbohydrate epitopes such as sialyl Lewis a and sialyl Lewis x expression<sup>41</sup> as well as through the interaction with ICAM-1.36 Itoh et al20 have studied the expression of MUC1, MUC2 and mucin carbohydrate antigens (Tn, sialyl-Tn, T, Lewis x, Lewis y and sialyl Lewis x-i) in normal, dysplastic and carcinoma of the pharynx and larynx; they found that only MUC1 was an effective marker of dysplasia and carcinoma in these localizations being Tn antigen expressed exclusively by carcinoma specimens; T antigen expression increases according with malignant progression. In the present approach, one of the most significant finding was that only one specimen failed to react with the MAbs generated against mucin and/or carbohydrate related antigens. It is interesting to point out that these carbohydrate antigens may be carried on by a variety of carrier molecules, e.g., lipids, glycoproteins and mucins; this fact may partly explain the reaction in specimens number 2 and 6 of carbohydrate moieties without mucin expression.

The coexpression of Lewis y and sialyl Lewis x in samples that were also reactive with anti Lewis x was also a remarkable finding taking into account that they are structures based on type 2 (Gal14GlcNAc-) carbohydrate sequence; Lewis y is the result of further fucosylation of the basic Lewis x antigen being sialyl Lewis x the consequence of the addition of sialyc acid to its terminal galactose. <sup>14</sup> On the other hand, Tn hapten is the first step of O-glycosylation of mucins such as MUC1 which make reasonable their co expression in some specimens; it is possible that in two samples which were reactive with Tn but did not show expression of MUC1, other glycoproteins and glycolipids could be Tn hapten possible carriers. <sup>10</sup>



**Figure 3.** Example of Western blotting analysis of a fraction obtained by density gradient centrifugation in guanidine/CsCl (fraction 7 from a density of 1.43 g/ml) as described in Methods. The fraction belonged to subcellular membranes from larynx carcinoma; the reaction with anti MUC1 C595 Mab is depicted. It shows the presence of two bands of a high-molecular-mass material of more than 200 kDa and some trace minor bands.

The isolation and further investigation of tumor associated antigens should be incremented in order to design directed therapies which can be useful as anticancer strategies. Different authors have employed with success similar procedures to isolate glycoproteins from diverse materials, 6 including mucins from tumor specimens; 1,40 we purified MUC1 through CSCl gradients and when fractions were analysed by Western-blotting with further incubation with anti-MUC1 C595 Mab, they revealed two bands which correspond to reduced mucin subunits; similar findings have been already identified in ENM belonging to breast carcinoma specimens<sup>7</sup> which were described by other authors. Furthermore, associated mucin carbohydrates have been detected such as Lewis x, Lewis y, sialyl Lewis x and Tn hapten; some of these were described linked to the protein core of mucins.<sup>44</sup>

In recent years, among prognostic factors determining the course of different tumor localizations, genetic alter98 CROCE et al

ations such as those affecting p53 have been the focus of considerable interest; mutations in p53 result in the synthesis of a protein with longer half-life, which is immunohistochemically detectable.<sup>17</sup> Its routine clinical application is a promising field since p53 overexpression and tumor chemosensitivity has been emphasised by Bradford et al.<sup>3</sup> We have included p53 which has been largely studied as a prognostic factor in laryngeal cancer with controversial significance about its reliability 5. We found that all tumor samples expressed p53 being sporadically found in normal tissues; this would be in coincidence with data involving this molecule with the progression of malignant tumors, although its exact role has not been yet elucidated. Previous studies<sup>26,29</sup> have shown associations between high levels of p53 expression and advanced disease stage as well as metastatic tumor dissemination, although p53 over expression has been identified as an early event in the development of carcinoma of the larynx.9 On the other hand, different efforts have been performed trying to establish p53 expression as a useful prognostic indicator but it does not appear to be of clinical value in this last aspect.37

Cytokeratins 8,18 and 19 that are normally expressed in head and neck tissue only in small amounts, become over-expressed in head and neck carcinoma; furthermore, cytokeratins 5/6 and 13 have been detected more frequently in metastatic squamous cell carcinoma than in the primary tumors. The anticytokeratin MAb employed in the present research stains cytokeratins 5, 6, 8, 17 and 19; hence it is not surprising that all tumor specimens were reactive being normal samples sporadically positive stained; these differences may reflect neoplasic phenotypic changes involved in invasive, migrating and proliferative activities.

In the present study, we have generated information on antigenic expression of carcinoma of the larynx that could be used for active and passive immunotherapy. An additional goal of our research was to perform a routine procedure in larynx carcinoma which covered different programmed studies, favouring the detection and isolation of MUC1 previously detected by immunohistochemistry.

## Acknowledgements

Thanks are expressed to Dr N. Hanai (Kyowa Hakko Tokyo Res. Lab., Tokyo, Japan) for the KM-93 and KM380 Mabs, to Prof. J. Taylor-Papadimitriou (The Imperial Cancer Research Fund, Guy Hospital, London, UK) for HMFG1 and HMFG2 MAbs, Dr P. Nielsen (School of Dentistry, Faculty of Health Sciences, Copenhagen, Denmark) for PANH2, PANH3 and PANH4 MAbs; to Dr E. Osinaga (Laboratorio de Oncología Básica, Facultad de Medicina, Montevideo, Uruguay) for the 83D4 Mab developed by Dr C Roseto (Université de Compiegne, France); to Dr CA Pereyra for providing patients' samples and to Dr. J Carri for encourage in photographical techniques.

### References

- 1.²Aksoy N, Corfield AP, Paraskeva C, et al. Human intestinal mucin MUC2 in in vitro two adenoma-carcinoma sequences originated from a single benign colonic adenoma cell line: In vitro evidence for the neotransformation. 5<sup>th</sup> International Workshop on Carcinoma Associated Mucins, Cambridge, UK, 1998
- 2. \*Barnd DL, Lan M, Metzgar R, et al: Specific, MHC-unrestricted recognition of tumor associated mucins by human cytotoxic T cells. Proc Natl Acad Sci USA; 86:7159-7163, 1989.
- 3. \*Bradford CR: Predictive factors in head and neck cancer. Hemotol Oncol Clin North Am 13:777-785, 1999.
- 4. \*Brown A, Feizi T Gooi HC: A monoclonal antibody against human colonic adenoma recognizes a difucosylated type-2 blood group chains. Biosci Rep 3:163-169, 1983.
- 5.2Chiesa F, Mauri S, Tradati N, et al: Surfing prognostic factors in head and neck cancer at the Millennium. Oral Oncol 35:590-596, 1999
- 6.2Creeth JM, Ramakrishnan Bhaskar K, et al: The separation and characterization of bronchial glycoproteins by density-gradient methods. Biochem J 167:557-569, 1977.
- 7.2Croce MV Price MR, Segal-Eiras A: Expression of monoclonal antibody-defined antigens in fractions isolated from human breast carcinoma and patients' serum. Cancer Immunol Immunother; 40:132-137, 1995.
- 8.<sup>2</sup>Croce MV Colussi AG, Price M, et al: A Immunohistopathological characterization of spontaneous metastases in a human lung mucoepidermoid adenocarcinoma (HLMC) xenograft. Pathol Oncol Res 4:259-266, 1998.
- 9. \*2Dolcetti R, Doglioni C, Maestro R, et al: A p53 over expression is an early event in the development of human squamous-cell carcinoma of the larynx: genetic and prognostic implications. Int J Cancer 52:178-182, 1992.
- 10.2 Federici MF, Kudryashov, Saigo PE, et al: Selection of carbohydrate antigens in human epithelial ovarian cancers for immunotherapy: serous and mucinous tumors exhibit distinctive patterns of expression. Int J Cancer 81:193-198, 1999.
- 11.2Feickert HJ, Anger BR, Cordon-Cardo, et al: Cell-surface antigens of human lung tumors detected by mouse monoclonal antibodies: definition of blood-group- and non-blood-group-related antigenic systems. Int J Cancer: 46:1007-1013, 1990.
- 12.2 Gendler SJ, Taylor-Papadimitriou J, Duhig T, et al: A highly immunogenic region of human polymorphic mucin expressed by carcinomas is made up of tandem repeats. J Biol Chem 263:12820-12823, 1988.
- 13.2Gendler SJ, Spicer AP: Epithelial mucin genes: Annu Rev Physiol 57:607-634, 1995.
- 14. Hakomori S, Nudelman E, Levery SB, et al: Novel fucolipids accumulating in human adenocarcinoma: I. Glycolipids with dior trifucosylated type 2 chain. J Biol Chem 259:4672-4680, 1984.
- 15.2 Hanai N, Shitara K, Yoshida H: Generation of monoclonal antibodies against human lung squamous cell carcinoma and adenocarcinoma using mice rendered tolerant to normal human lung. Cancer Res 46:4438-4443, 1986.
- 16.ºHilkens J, Ligtenberg JL, Vos HL, et al. Cell membrane-associated mucins and their adhesion-modulating property. Trends Biochem Sci 17:359-363. 1992.
- 17. Hurlimann J, Saraga EP. Expression of p53 protein in gastric carcinomas. Am J Surg Pathol 18:1247-1253, 1994.
- 18. International Union Against Cancer (UICC) Classification of Malignant Tumors. Ed. L.H. Sobin and Ch. Wittekind, 5th edition, New York, 1997.

- 19. Ioannides CG, Fisk B, Jerome KR, Irimura T. Cytotoxic T cells from ovarian malignant tumors can recognize polymorphic epithelial mucin core peptides. J Immunol 151: 3693-3703, 1993.
- 20.2 Itoh T. Vonezawa S, Nemoto M, et al.: Expression of mucin antigen and Lewis x-related antigens in carcinomas and dysplasia of the pharynx and larynx. Pathol Int 46: 646-655, 1996.
- 21. 2Ierome KR, Barnd DL, Bendt KM: Cytotoxic T-lymphocytes derived from patients with breast adenocarcinoma recognize an epitope present on the protein core of a mucin molecule preferentially expressed by malignant cells. Cancer Res 5:2908-2916, 1991.
- 22. Johnson NW: Orofacial neoplasms: global epidemiology, risk factors and recommendations for research. Int Dental J 41:365, 1991.
- 23. \*Kotera Y, Darrell Fontenot J, Pecher G, et al. Humoral immunity against a tandem repeat epitope of human mucin MUC1 in sera from breast, pancreatic and colon cancer patients. Cancer Res 54:2856-2860, 1994.
- 24. Laemmli UK: Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature (Lond) 227:680-681, 1980.
- 25.2McDonald LA, Walker DM, Gibbins JR: Cervical lymph node involvement in head and neck cancer detectable as expression of a spliced transcript of type II keratin K5. Oral Oncol 34:276-283, 1998.
- 26.²Nees M, Homann N, Discher H, Andl T, et al: Expression of mutated p53 occurs in tumor-distant epithelia of head and neck cancer patient: a possible molecular basis for the development of multiple tumors. Cancer Res 53:4189-4196, 1993.
- 27. Nielsen PA, Mandel U, Therkildsen MH, et al: Differential expression of human high-molecular-weight salivary mucin (MG1) and low-molecular-weight salivary mucin (MG2). J.Dent Res 75:1820-1826, 1996.
- 28. Nielsen PA, Mandel U, Therkildsen MH, et al: Loss of a novel mucin-like epithelial glycoprotein in oral and cervical squamous cell carcinoma. Cancer Res 57:634-640, 1997.
- 29. Nylander K, Stenling R, Gustafsson H, et al: p53 expression and cell proliferation in squamous cell carcinomas of the head and neck. Cancer 75:87-93, 1994.
- 30.2Pancino GF, Osinaga E, Vorauher W et al: Production of a monoclonal antibody as immunohistochemical marker on paraffin embedded tissues using a new immunization method. Hybridoma 9:389-395, 1990.
- 31.2Parkin DM, Pisani P, Ferlay J: Estimates of the worldwide incidence of eighteen major cancers in 1985. Int J Cancer 54:594-599, 1993.
- 32.2 Price MR, Edwards S, Owainati A, et al: Multiple epitopes on a human breast carcinoma associated antigen. Int J Cancer 36:567-574, 1985.
- 33.2Price MR, Pugh J, Hudecz F, et al: Immunological and structural features of the protein core of human polymorphic epithelial mucin. Mol Immunol 27:795-801, 1990.

- 34. Quinlan DC, Davidson AG, Summers CL, et al: Accumulation of p53 protein correlates with a poor prognosis in human lung cancer. Cancer Res 52:4828-4831, 1992.
- 35.2 Ravindranath NH, Nishimoto K, Chu K, et al: Density of cell surface expression of sialyl Lewis antigens and complement restriction factors in oral squamous cell carcinoma. Proc Am Ass for Cancer Res 40:357, 1999.
- 36.2Regimbald LH, Pilarski LM, Longenecker BM: The breast mucin MUC1 as a novel adhesion ligand for endothelial intercellular adhesion molecule 1 in breast cancer. Cancer Res 56:4244-4249, 1996.
- 37.2 Rowley H, Poland NJ, Helliwell TR, et al: P53 protein expression in tumors from head and subsites, larynx and hypopharynx and differences in relationship to survival. Clin Otolaryngol 23:57-62, 1998.
- 38.2 Rughetti A, Turchi V Apollonj Ghetti C: Human B-cell immune response to the polymorphic epithelial mucin. Cancer Res 53:2457-2459, 1993.
- 39.2 Suo Z, Holm R, Nesland JM: Squamous cell carcinomas. An immunohistochemical study of cytokeratins and involucrin in primary and metastatic tumors. Histopathol 23:45-54, 1993.
- 40.2 Switacheva N, Davies JR, Lesuffleur T: Characterization of mucins produced by human HT29-MTX cell line. 5th International Workshop on Carcinoma Associated Mucins Cambridge, UK. 1998.
- 41.2 Takada A, Ohmori K, Yoneda T: Contribution of carbohydrate antigens sialyl Lewis a and sialyl Lewis x to adhesion of human cancer cells to vascular endothelium. Cancer Res 53:354-361, 1993.
- 42. Taylor KL, Mall AS, Barnard RA: Immunohistochemical detection of gastric mucin in normal and disease states. Oncol Res, 10:465-473, 1998.
- 43. \*\*Taylor-Papadimitriou J, Peterson J, Arklie J: Monoclonal antibodies to epithelium-specific components of the human milk fat globule membrane: production and reaction with cells in culture. Int J Cancer 28:17-21, 1981.
- 44. Taylor-Papadimitriou J, Finn O: Biology, biochemistry and immunology of carcinoma associated mucins. Immunol Today 18:105-107, 1997.
- 45. Thornton DJ, Devine PL, Hanski Ch, et al: Identification of two major populations of mucins in respiratory secretions. Am J Respir Crit Care Med 150:823-832, 1994.
- 46.2 Towbin H, Staehelin T Gordon J: Electrophoretic transfer of proteins from polyacrilamide gels to nitrocellulose sheets: procedures and some applications. Proc Natl Acad Sci USA 76:4350-4354, 1979
- 47.2 Whitehouse C, Burchell J, Gschmeissner S: A transfected sialyltransferase that is elevated in breast cancer and localizes to the medial/trans Golgi apparatus inhibits the development of core-2-based O-glycans. J Cell Biol 137:1229-1241, 1997.